



ENTOMO TAXONOMY LAB [ETL]





CHRIST COLLEGE (AUTONOMOUS) IRINJALAKUDA



IRINS Profile

DR. ABHILASH PETER ASSISTANT PROFESSOR & RESEARCH SUPERVISOR ZOOLOGY DEPARTMENT Entomo Taxonomy Lab was established on November 2020 with an aim to inculcate entomo research in young aspirants. ETL mainly focus on taxonomic and life cycle studies of moths (Lepidoptera: Heterocera) which are sister group of butterflies. It also give prime importance to research on the association of natural enemies particularly parasitic hymenopterans (Hymenoptera) with moth's developmental stages (egg, larva and pupa) and thereby its application as biological control agents in agroecosystems.

Research Area

- Taxonomy studies on Parasitic Hymenoptera.
- Taxonomic and diversity studies on moths.
- Life cycle studies on moths.
- Published many new species of parasitic hymenoptera (click IRINS)
- Published new host plant records for moths.



Research Students





Mr. Adarsh P. K.

Research Area: Moths of Superfamily Noctuoidea.

Fellowship: UGC-JRF



Mrs. Aiswarya N.

Research Area: Moths and associated parasitic hymenopterans on developmental

stages of moths.

Fellowship: UGC-JRF

Entomo Taxonomy Lab

Lab Facility



Labomed Microscope for identification of specimens



Ample working space





Spreading board for proper wing spreading of moths.



Insect Box- Used to preserve dried moth specimens in air tight box to avoid damage.



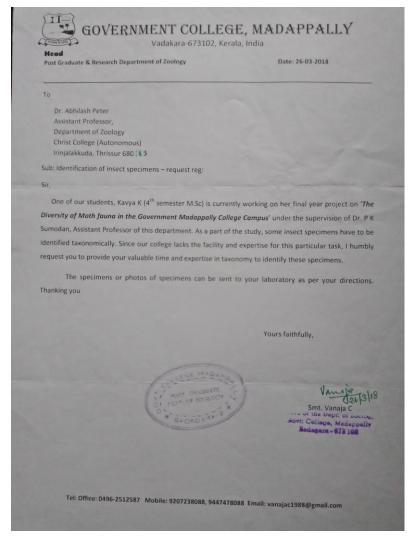
Rearing- Plastic jars and rearing cages are used to study developmental stages of moths and associated parasitic hymenopterans

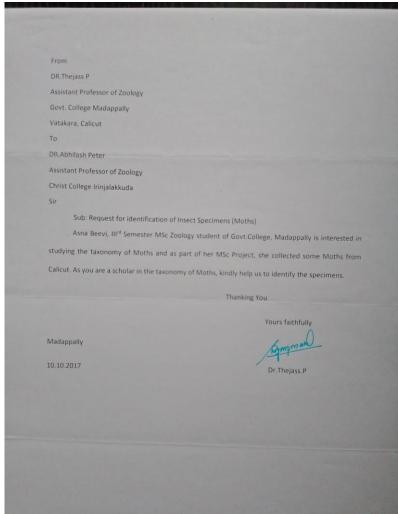


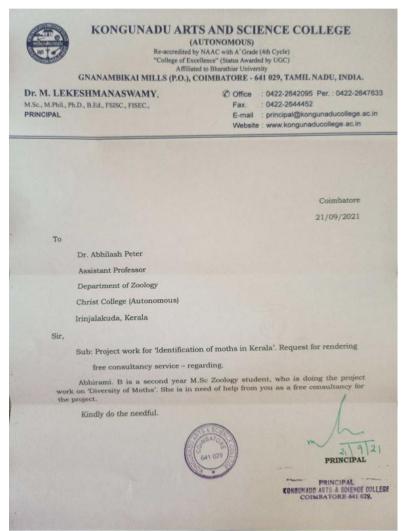


Services

- Identification of insects particularly moths of BSc students UG project on request.
- Cataloguing of moths as part of MSc dissertation of PG students from various colleges on request.











Post Graduate and Research Department of Zoology

GOVERNMENT COLLEGE, MADAPPALLY

(Accredited at A level (third cycle) by NAAC)
VATAKARA, KOZHIKODE-673102
KERALA, INDIA

Dr.Thejass P Assistant Professor thejassp@gmail.com 09947361321

To

Dr. Abhilash Peter

Assistant Professor of Zoology

Christ College Irinjalakkuda

Thrissur

Sir,

Sub: Identification of Moths for BSc Project.

Final year BSc students of this college has been doing a project related to diversity of moths as part of their curriculum under my supervision. As you are an expert in studies related to Moths, I humbly request your help for the identification of Moth specimens collected by my students.

Thanking You

Yours sincerely



Date: 15 March 2022

Services

A Partial List of Moths (Leidoptera: Heterocera) from Kozhikode District, Kerala, India

Abhilash Peter1 and Adarsh P. K.2

Abstract:

The present paper deals with the moths collected from different localities of Kozhikode district, Kerala. The information presented in this paper is based on the insect surveys conducted from 01-07 2004 to 28-02-2005. Of the total 300 specimens collected, 139 species belonging to 108 genera under 18 families were identified to the species level. Family Erebidae was found to be dominant in terms of number of species when compared to the other families. An updated systematic account is provided for all the identified specimens.

Key words: list, moths, Kozhikode, Kerala,

Introduction

Kozhikode district lies between 11° 08'N and 11° 50'N latitudes and 75°30'E and 76°8'E longitudes. District is bor-

- 1 Assistant Professor, Department of Zoology, Christ College (Autonomous), Irinialakuda-680125
- 2 Department of Zoology, Christ College (Autonomous), Irinjalakuda-680125

Recherche - A Digest of Multidisciplinary Explorations (ISBN 978-93-91343-03-3) | 219



Publications

dered by Kannur district to the North, Malappuram to the South, Wayanad to the East and Arabian Sea to the West. The region receives an appreciable amount of rainfall every year (South-West and North-East Monsoon). The district is blessed with lush green vegetation in the Western Ghat part comprising trees, shrubs herbs, climbers etc. and forms a hiding place for many animal species.

Lepidoptera includes butterflies and their sister group, moths. Though harmless, moths are serious pests of many agricultural crops and commercial plants. This study is an attempt to know the species of moths from Kozhikode district.

Methodology

Moths were collected from different localities of Kozhikode district which includes plains, hilly areas and forests during the period 01-07 2004 to 01-02-2005. Live specimens from Kakkayam forest, Chalappuram, Janakikkadu, Thushsragiri, Nadakkavu, East Hill, and Chalappuram were collected mainly by hand picking method. Those specimens attracted to light were collected by using an insect net. A light trap was operated overnight during the mothing season and occasional collection was also done from other areas of Kozhikode district. Live specimens were killed in a killing jar filled with ethyl acetate vapours. The specimens were then dried and preserved in air tight insect boxes with appropriate data labels.

Identification of moths was done with the help of relevant literatures like, Hampson (1892-96), Bell and Scott (1937, Holloway (1983-2005), Barlow and D'Abrera (1982), Robinson et al. (1994), Kendrick (2002, 2004) and Pittaway and Kitching (2004). The classification followed here is based on the literature published by Nieukerken et al. (2011) and Zahiri et al. (2012). The specimens were identified using Leica APO Stereozoom microscope. A digital camera Canon A620 was used for taking the photos of moths.



Family: Erebidae

Subfamily: Calpinae Eudocima phalonia Clerck (Fig.47,53) Eudocima hypermnestra (Cramer) (Fig.33) Eudocima materna Linnaeus Eudocima homaena (Hubner) Phyllodes consobrina Westwood Oraesia emarginata (Fabricius) Achaea ianatha (Linnaeus) Subfamily: Boletobinae

Lopharthrum comprimens Walker (Fig.25) Subfamily: Scoliopteryginae Anomis flava Fabricius

Subfamily: Pangraptinae

Egnasia accingalis Walker

Egnasia ephyrodalis Walker (Fig.2) Subfamily: Erebinae

Ischyja manlia (Cramer) (Fig.28) Dysgonia stuposa Fabricius Bastilla crameri Moore (Fig.14) Grammodes geometrica (Fabricius) Hulodes caranea Cramer Ercheia cyllaria Carmer (Fig.17) Oxyodes scrobiculata Fabricius Erebus hieroglyphica Drury (Fig.49) Thyas coronata Fabricius Thyas honesta Hubner (Fig.18) Lygniodes vampyrus (Fabricius) Artena submira Walker Trygonodes hyppasia Crame Erebus macrops Linnaeus (Fig.9) Ericeia inangulata (Guenee) Dierna patibulum Fabricius Avatha bubo (Geyer) (Fig.11) Serrodes campana Guenee Sphingomorpha chlorea (Cramer) Spirama retorta Clerck (Fig.5)

Mocis undata Fabricius (Fig.45) Mocis undata Fabricius (Fig.4: Subfamily: Aganainae Asota producta Butler (Fig.6) Asota caricae Fabricius

Asota heliconia (Linnaeus) Asota plana (Walker) Asota ficus Fabricius Asota ficus Fabricius Neochera inops (Walker) (Fig.1)

Family: Lasiocampidae Subfamily: Lasiocampinae

amily: Sphingidae Subfamily: Macroglossinae

Macroglossum insipida Butler Macroglossum gyrans Walker Acosmeryx anceus subdentata Rothschile & Jordan (Fig.3)

Angonyx testacea (Walker)

Angonyx testacea (Walker)

Hippotion celerio (Linnaeus)

Hippotion boerhavia (Fabricius) (Fig.24)

Theretra lycetus Cramer (Fig.41)

Theretra oldenlandae (Fabricius) (Fig.31) Theretra pallicosta (Walker) (Fig.50) Theretra silhetensis Walker (Fig.40) Theretra latreillii (Macleay) Theretra nessus (Drury) Theretra gnoma (Fabricius) Theretra alecto (Linnaeus) Pergesa acteus (Cramer) (Fig.46) Nephele hespera (Fabricius) (Fig.26)

Marumba dyras (Walker) (Fig.15) Subfamily: Sphinginae Acherontia styx Westwood

Acherontia lachesis (Fabricius) (Fig.19) Psilogramma sp. (menephron agg.) (Fig. 34 Agrius convolvuli Linnaeus (Fig.32) Subfamily: Smerinthinae

Amplypterus panopus Cramer (Fig.27) Daphnis nerii (Linnaeus) (Fig.37) Ambulyx belli (Jordan) (Fig.23)

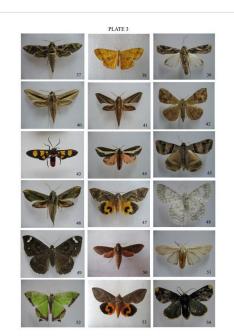
Subfamily: Eupterotinae Eupterote undata Blanchard Eupterote mollifera Walker

Family: Lasiocampidae Subfamily: Lasiocampinae Trabala vishnou Lefebvre (Fig.13) Family: Uraniidae

Subfamily: Epipleminae Orudiza protheclaria Walker Epiplema irrorata (Moore)

Subfamily: Microniinae

Micronia aculeata Guenee Family: Geometridae



UTTAR PRADESH JOURNAL OF ZOOLOGY

43(9): 76-92, 2022 ISSN: 0256-971X (P)



A PRELIMINARY CHECKLIST OF HYMENOPTERAN LARVAL AND PUPAL PARASITOIDS OF MOTHS (LEPIDOPTERA: HETEROCERA) FROM INDIA

AISWARYA NANDALAN 20 AND ABHILASH PETER 200

^a Department of Zoology, Christ College (Autonomous), Irinjalakuda, Thrissur, Kerala - 680125, India.

AUTHORS' CONTRIBUTIONS

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

- (1) Dr. Juan Carlos Troiano, University of Buenos Aires, Argentina.
- Reviewers:
 (1) Mahendran B, Indian Council of Agricultural Research (ICAR), India.
- (2) Ibrahim El-Sayed Shehata, National Research Centre, Egypt.
- (3) Carlos Henrique Marchiori, Instituto Federal Goiano, Brazil.

Received: 11 March 2022 Accepted: 22 May 2022 Published: 28 May 2022

Review Article

ABSTRACT

The present paper, based on the study of published literatures, provides an updated checklist of 54 species of pupal and 197 species of larval hymenopteran parasitoids associated with the larva and pupa of moths from India. Though the literatures pertaining to the above research work are available in various published journals and books, a compiled list is not so far published from India. The present paper also included the systematic status of both host (moth) and its natural enemies (hymenopteran parasitoids), which would be helpful for biological control workers in future.

Keywords: Larval; pupal parasitoids; moths; heterocera; India.

1. INTRODUCTION

Lepidoptera, one of the largest insect orders after beetles, comprises more than 160,000 described species globally. According to van Nieukerken et al. [1], there are 15,578 described genera and 157,424 species worldwide. They are treated as sister group of butterflies and have an unwelcome impact on crops and other economically important flora. Many moth

agroecosystems. The conventional way of controlling these pests is by the use of insecticides or other chemicals which could bring about a non reversible damage to the genetic machinery of other fauna [2-

To reduce crop production losses and to diminish insecticide use, entomologists provided an ecofriendly method namely, biocontrol agents in which species are polyphagous and are major pests of the natural enemies of the pests are used to control its

[&]quot;Senior Research Fellow,

Assistant Professor;

^{*}Corresponding author: Email: abhilashpeter@gmail.com;